

# **Molecular virology - Laboratory exercises -**

## **SDS-PAGE ANALYSIS OF VIRAL PROTEINS**

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## CHARACTERISTICS OF POLYACRYLAMIDE GELS:

- Synthetic polymer
- Chemically inert
- Irreversible chemical polymerisation
- Pore size is well defined – good resolution
- Good mechanical properties
- Applicable for analysis of proteins and nucleic acids

<u>% Acrylamide</u>	<u>MW Range (kDa)</u>
7	50 – 500
10	20 – 300
12	10 – 200
15	3 - 100

### Advantages of PAGE in NA analysis

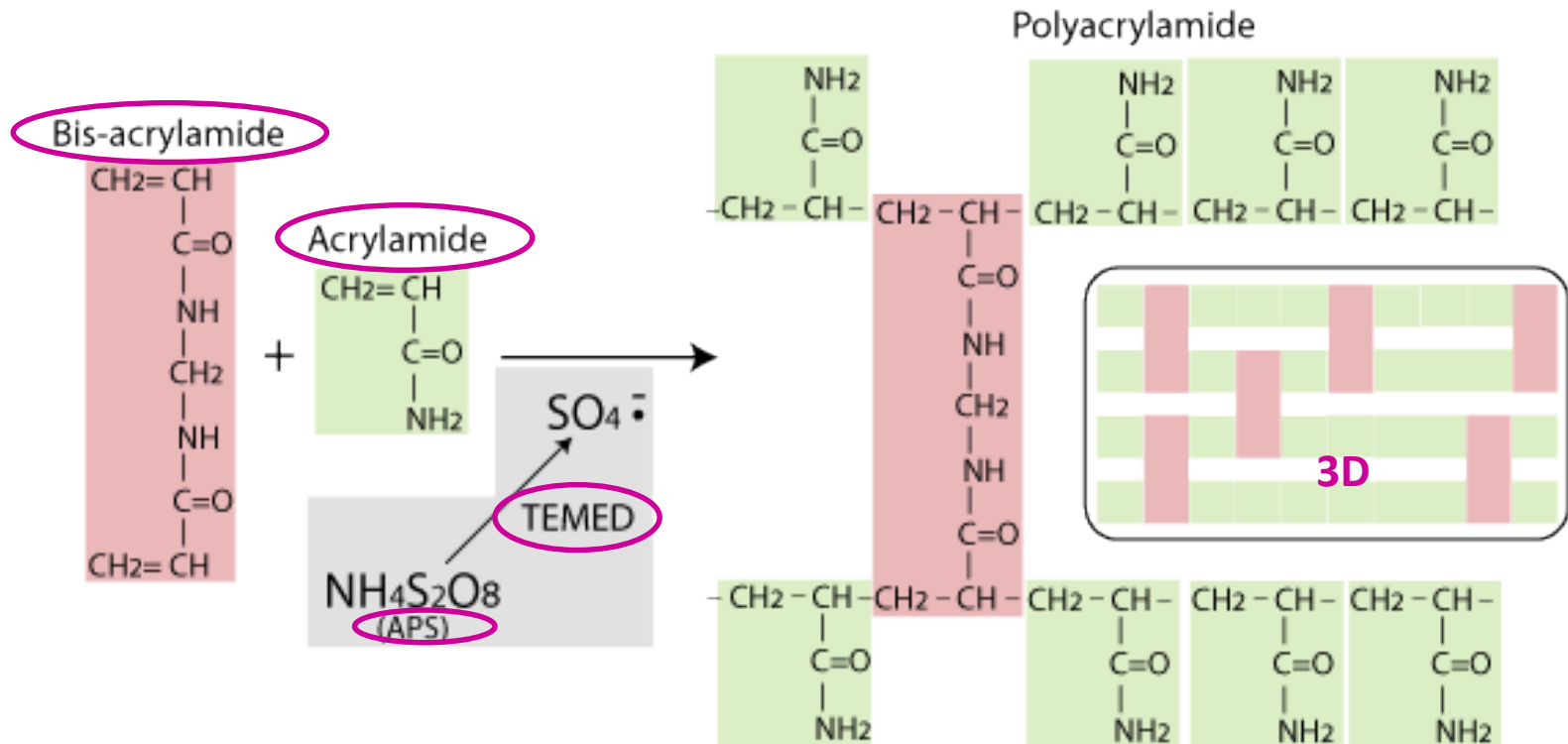
- Higher sample load per well
- Better NA purity for further analysis
- 0.2% resolution

## Composition of polyacrylamide gel (10 mL, 8%):

6.63 mL EF-buffer  
1.93 mL **AA** (40%)  
1.33 mL **BIS** (2%)  
100  $\mu$ L **APS** (10%)  
10  $\mu$ L **TEMED**

EF buffer = electrophoresis buffer  
AA = acrylamide  
BIS = bis-acrylamide  
APS = ammonium persulfate  
TEMED = tetramethylethylenediamine

polymerisation for **15 min** at room temperature

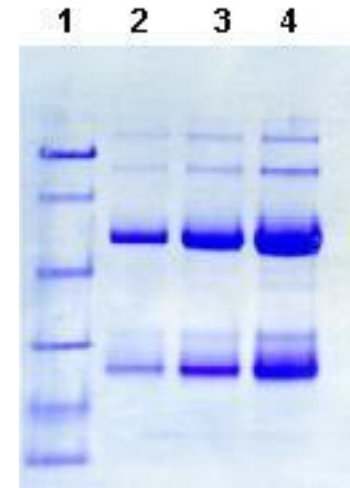


**APS** – source of free radicals for polymerisation

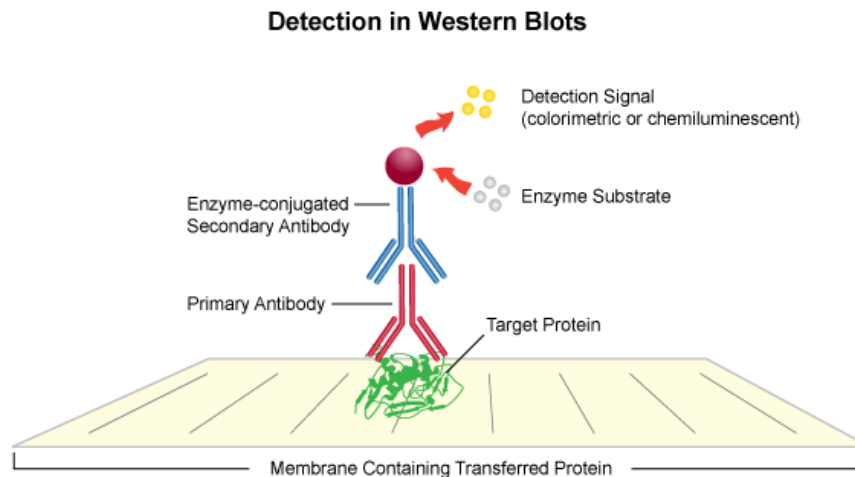
**TEMED** – catalyst, induces the production of free radicals

## APPLICATION IN VIROLOGY:

- **Identification** of specific viral proteins (structural or functional, e.g. reverse transcriptase)



➔ often followed by **western blot**  
e.g. detection of specific antibodies in serum (HIV)



**Diagram 2:** Illustration of detection in Western Blots.

# SDS-PAGE vs NATIVE PAGE

Electrophoretic mobility depends on:

**charge** (dependent on buffer pH)

**shape**

**size**

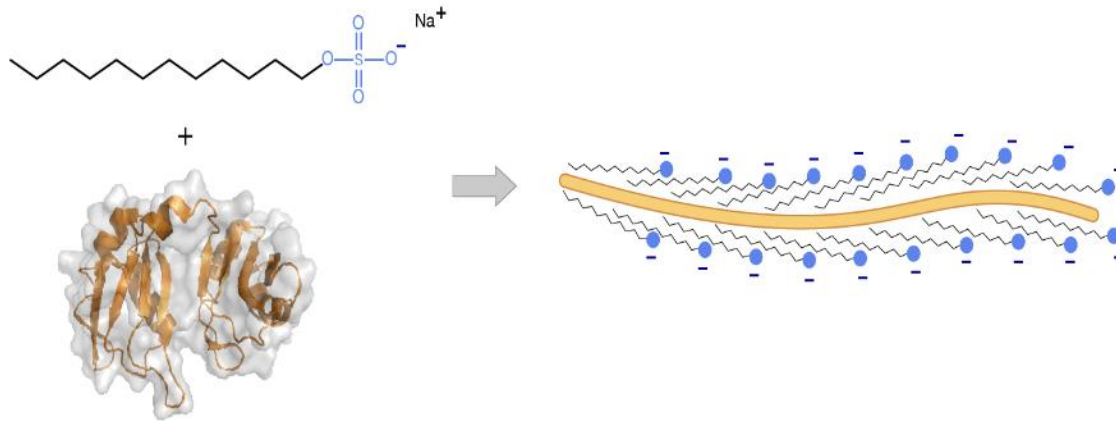
**mass** of molecules

**Sodium dodecyl sulphate (SDS)** or **sodium lauryl sulphate (SLS)** – anionic detergent; acts as a denaturing agent and a surfactant, binding to the denatured protein and **masking** its charge

1 g of protein binds 1.4 g of SDS

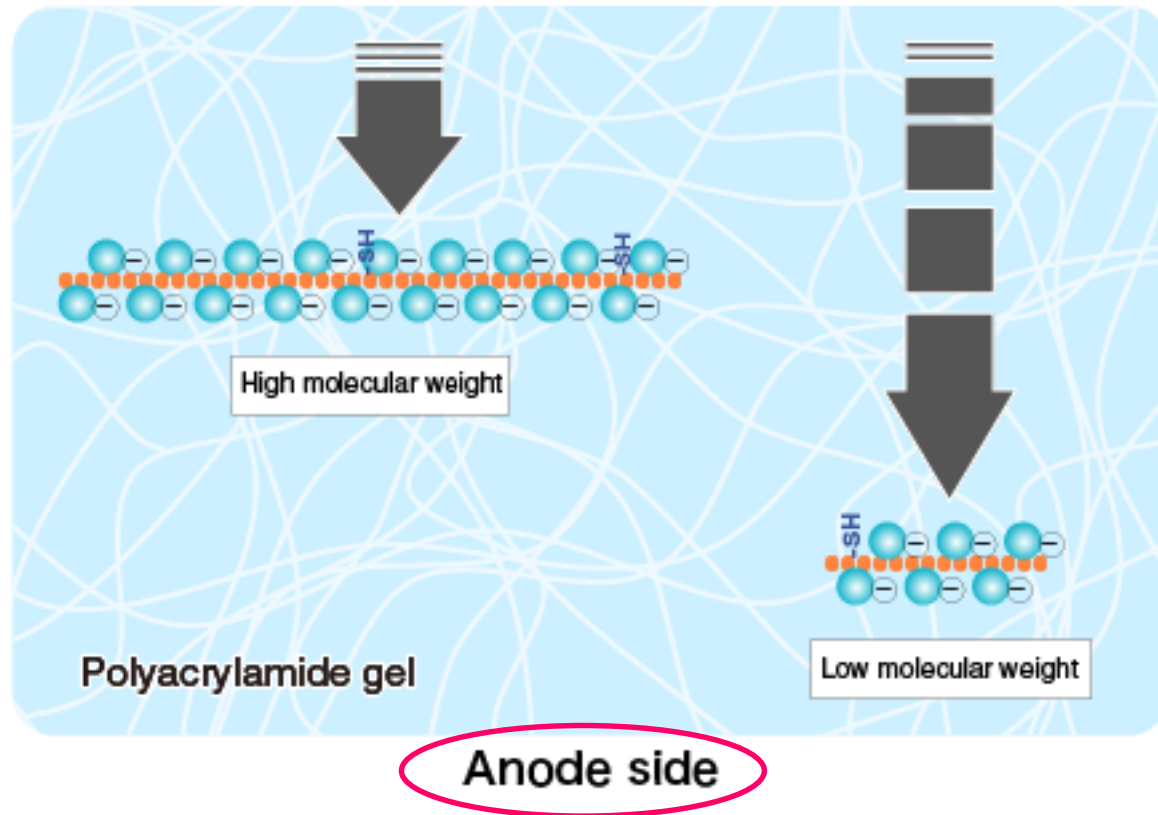


**constant charge-to-mass ratio**



Proteins are separated based on their polypeptide chain length by electrophoresis in a polyacrylamide gel with an appropriate mesh size.

Cathode side



Separation based on difference in molecular mass

## PROCEDURE:

- preparation of the gel
- preparation of samples
- electrophoresis
- staining

## SAMPLES:

- freshly purified TMV, negative control, lyophilised TMV-protein
- freshly purified : PVX, CMV, TBSV...
- Protein marker 10-250 kDa (New England Biolabs)

## Composition of polyacrylamide gel (10 mL, 8%):

6.63 mL EF-buffer  
1.93 mL AA (40%)  
1.33 mL BIS (2%)  
100  $\mu$ L APS (10%)  
10  $\mu$ L TEMED

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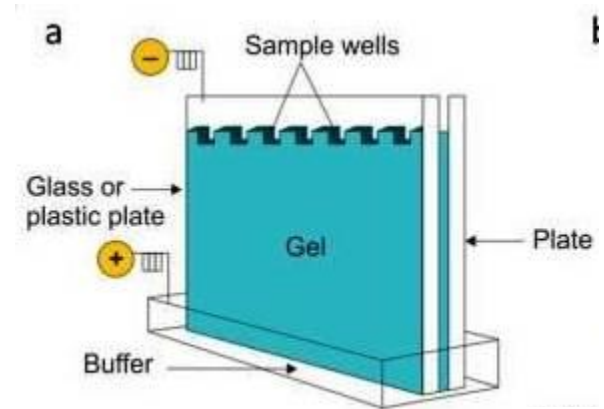
Polymerisation for **15 min** at room temperature

## Composition of electrophoretic buffer (1 L, 10x concentrated, pH 8.3):

30.3 g TRIS  
144 g glycine  
10 g SDS

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fill up to 1 L with deionised water





## Composition of Laemmli hydrolysis buffer (2x conc., 100 mL):

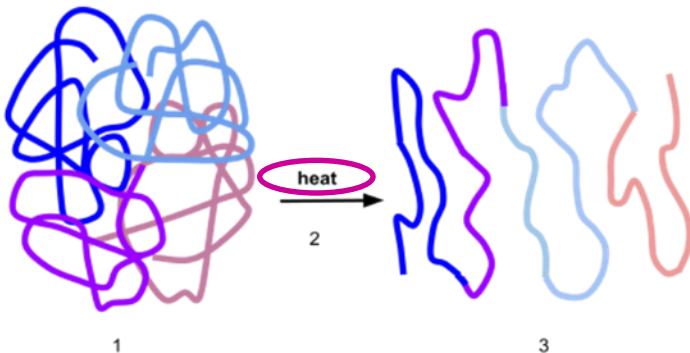
12 mL Tris-HCl (1M, pH 6.8)  
40 mL SDS (10%)  
20 mL glycerol  
10 mL  $\beta$ -mercaptoethanol  
0.01 g bromphenol blue (BPB)

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fill to 100 mL with deionised water

## Sample hydrolysis:

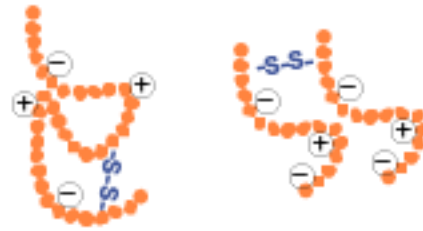
15  $\mu$ l sample  
15  $\mu$ l Laemmli hydrolysis buffer

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heat for **5 min at 95°C** and put on **ice**



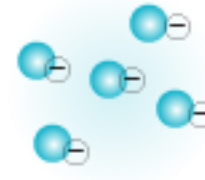
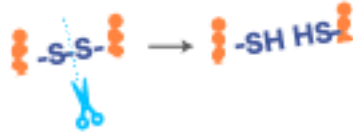
**SDS,  $\beta$ -mercaptoethanol and heat** – protein denaturation  
 **$\beta$ -mercaptoethanol** – reduces disulphide bonds  
**glycerol** – increases sample density

Proteins  
Folded with positive  
and negative charges



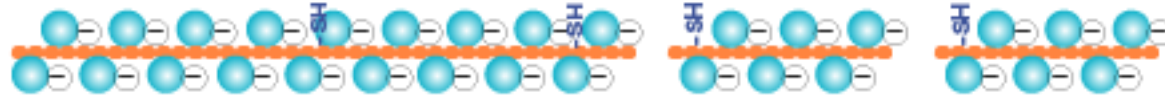
-S-S- : Disulfide bond

Reduced by 2-mercaptoethanol  
(disulfide bonds are reduced)

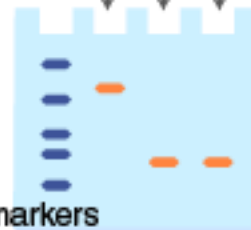


SDS with negative charge

Unfolded to a linear structure with negative charge proportional  
to the polypeptide chain length



Molecular weight markers



## Sample loading:

- protein marker – 7  $\mu\text{L}$  per well (*ready to use*)
- virus samples – 20  $\mu\text{L}$  per well

**60-90 min at 70 V**

## Composition of staining solution:

0.25% Coomassie Brilliant Blue

50% methanol

10% acetic acid

40% deionised water

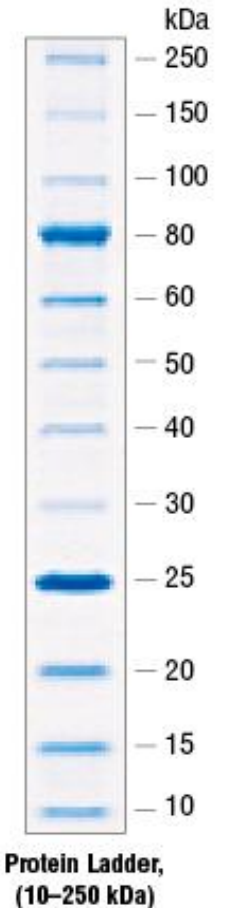
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incubate the gel for **10 min**

rinse with deionised water 2x

boil briefly in a microwave oven 2x

rinse with deionised water 2x

leave to de-stain on orbital shaker overnight



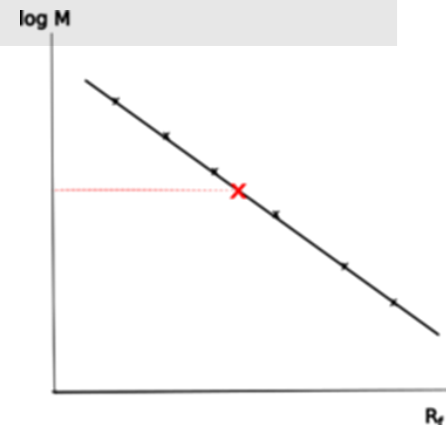
Alternative staining methods:

- silver – best sensitivity
- amido-black...

## EXERCISE:

protein	Molecular mass (Da)	Logarithm of molecular mass	Electrophoretic mobility (mm)
albumin	66000	4.81954	3
ovalbumin	45000	4.65321	11
pepsin	34700	4.53237	15
trypsin	24000	4.38021	22
$\beta$ - lactoglobulin	18400	4.25624	28
lysozyme	14300	4.14706	35

Protein X has electrophoretic mobility of **7.5**.  
Calculate its molecular mass!



## TASK:

Study all materials of virtual laboratory on link:

<http://vlab.amrita.edu/?sub=3&brch=186&sim=319&cnt=1>

and short video on apparatus assembly and gel casting:

[https://www.youtube.com/watch?v=EDi\\_n\\_0NiF4](https://www.youtube.com/watch?v=EDi_n_0NiF4)