

INTRODUCTION

Antimicrobial properties of silver and enhanced reactivity when applied in the nanoparticle form (AgNPs) has led to their increasing utilisation in consumer products. However, their increasing release into water or soil represents a potential environmental hazard. AgNPs can impose detrimental effects on plants, mainly through excess generation of reactive oxygen species (ROS), leading to induction of oxidative stress¹. In this work, detached roots of *in vitro* grown tobacco (*Nicotiana tabacum*) plants were exposed to AgNPs stabilised with cetyltrimethylammonium bromide (CTAB) or polyvinylpyrrolidone (PVP) coating or to ionic silver (AgNO₃), applied in the same concentration (100 µmol/L) for 24 h. The aim was to investigate *in situ* early physiological responses that are the first signs of stressful conditions. Generation of ROS in root cells was monitored by confocal laser scanning microscopy, using highly sensitive and specific fluorescent probes: dihydroethidium (DHE) to detect O₂⁻ and 2',7'-dichlorofluorescein-diacetate (H₂DCF-DA) to detect H₂O₂². Propidium iodide was used as a counterstain to verify cell viability³.

MATERIALS AND METHODS



RESULTS

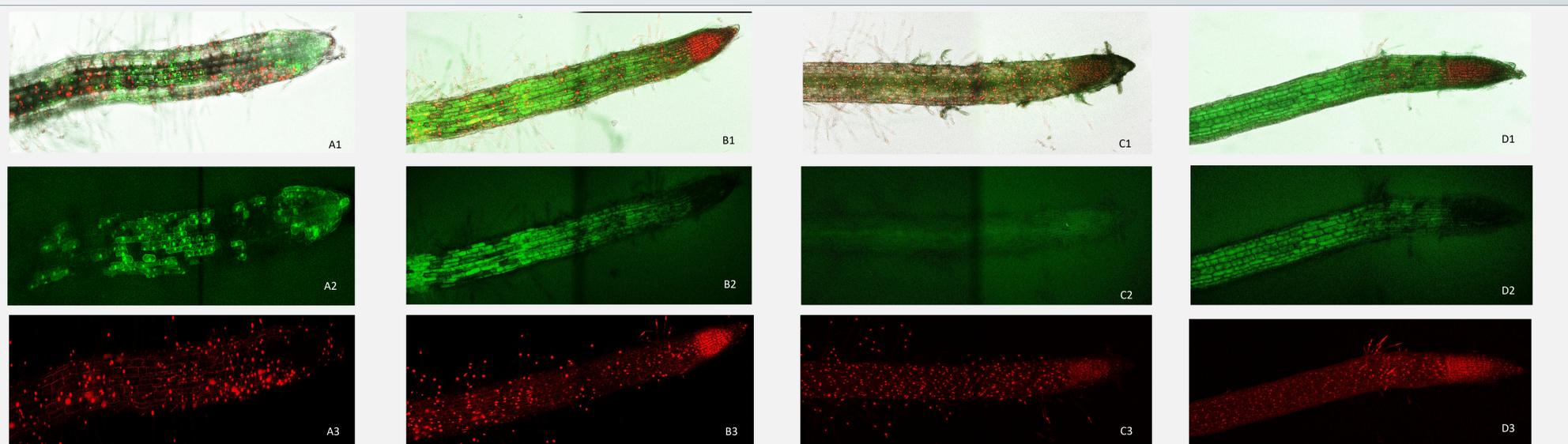


Figure 1. Confocal imaging of *N. tabacum* roots stained with H₂DCF-DA that labels H₂O₂ to give a green fluorescent product, after a 24 h incubation with specified treatments: (A1-3) control, untreated cells; (B1-3) cells treated with 100 µM AgNP-PVP; (C1-3) 100 µM AgNP-CTAB; (D1-3) 100 µM AgNO₃ solution. Red fluorescence corresponds to PI staining that labels nuclei of dead cells. All images represent maximum intensity projection of Z-stack of between 30 to 40 images (Z-step: 0,03 µm, magnification: 20X). Images A1, B1, C1 and D1 represent an overlay of brightfield channel, green and red fluorescence channels; whereas A2, B2, C2 and D2 represent only the green fluorescent channel (showing the levels of H₂O₂ in the cells) and A3, B3, C3 and D3 represent only the red fluorescent channel (showing the amount of dead cells stained red).

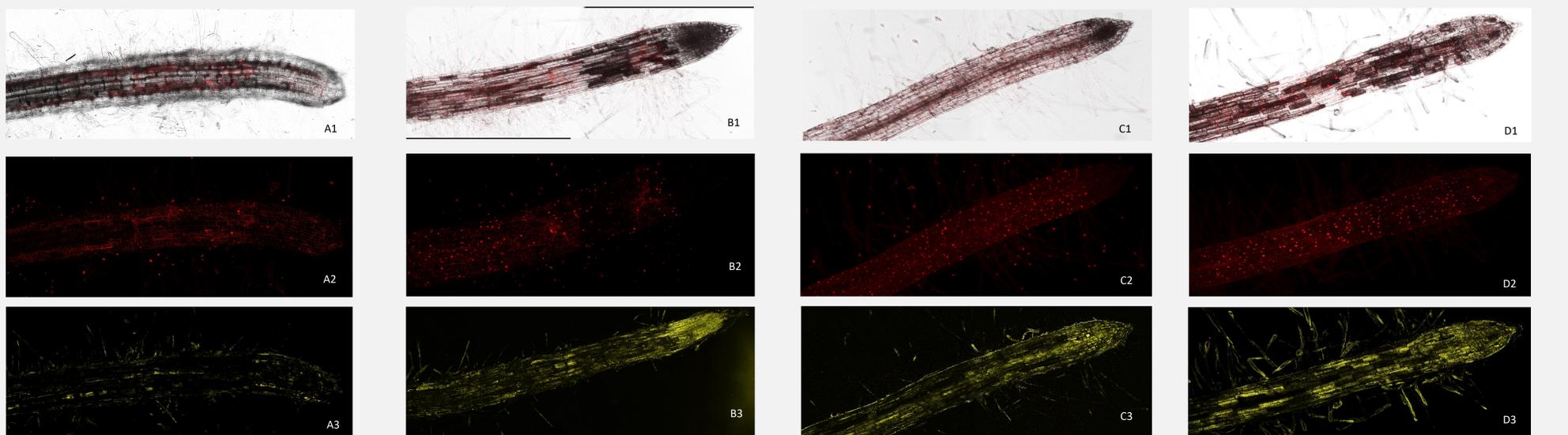


Figure 2. Confocal imaging of *N. tabacum* roots stained with DHE that labels O₂⁻ to give a red fluorescent product, after a 24 h incubation with specified treatments: (A1-3) control, untreated cells; (B1-3) cells treated with 100 µM AgNP-PVP; (C1-3) 100 µM AgNP-CTAB; (D1-3) 100 µM AgNO₃ solution. All images represent maximum intensity projection of Z-stack of between 30 to 40 images (Z-step: 0,03 µm, magnification: 20X). Images A1, B1, C1 and D1 represent an overlay of brightfield channel, red fluorescence channel and reflection (476 nm) channel; whereas A2, B2, C2 and D2 represent only the red fluorescent channel (showing the levels of O₂⁻ in the cells) and A3, B3, C3 and D3 represent only the reflection channel (where the yellow coloured areas most likely represent AgNPs).

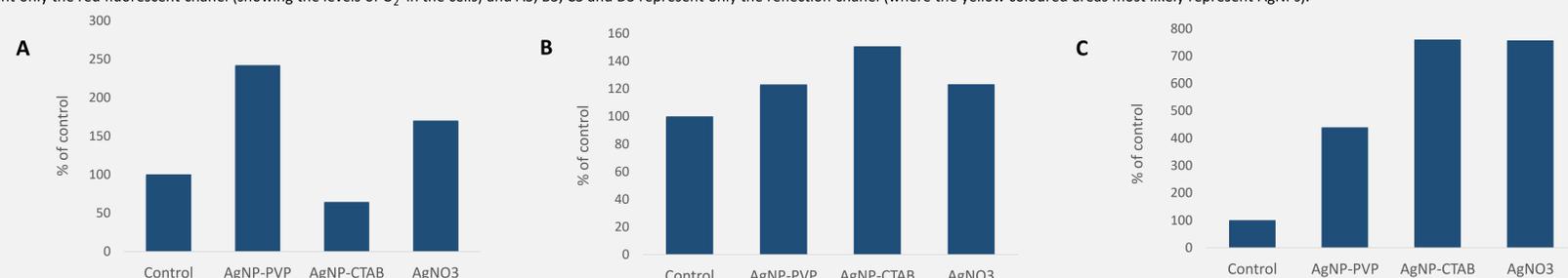


Figure 3. Graphs represent the mean pixel density evaluated from the maximum projection image of roots stained with (A) H₂DCF-DA; (B) PI; (C) DHE, after incubation with specified treatments. The results are expressed as % of control, where the control represents 100 %.

REFERENCES

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CONCLUSIONS

- Increased amount of H₂O₂ and O₂⁻ was observed in roots exposed to either AgNPs or AgNO₃ comparing to non-exposed, control roots, which was correlated to an increased percentage of dead cells; while the changes were most prominent after AgNP-CTAB treatment. Tissue necrosis was predominantly observed in root tips.
- Additionally, AgNPs were visualized and were mostly found attached to the root surface and on roots hairs, and their accumulation could be linked to excess generation of ROS.
- The results show that AgNPs not only compromised cellular redox-homeostasis but possibly caused cell necrosis, while different level of impact was observed depending on the form of applied silver and possibly on the intrinsic properties of AgNP coatings.